

5. Please use a standard 10 µl pipette to load your sample. Please load the sample by inserting the pipette tip vertically into the well. Maximal volume per well is 60 µl.



### 6. Electrophoresis Condition

Voltage	Starting current	Finished current	Run Time per Gel*
140 V	75 - 100 mA	30 - 50 mA	45 - 55 min

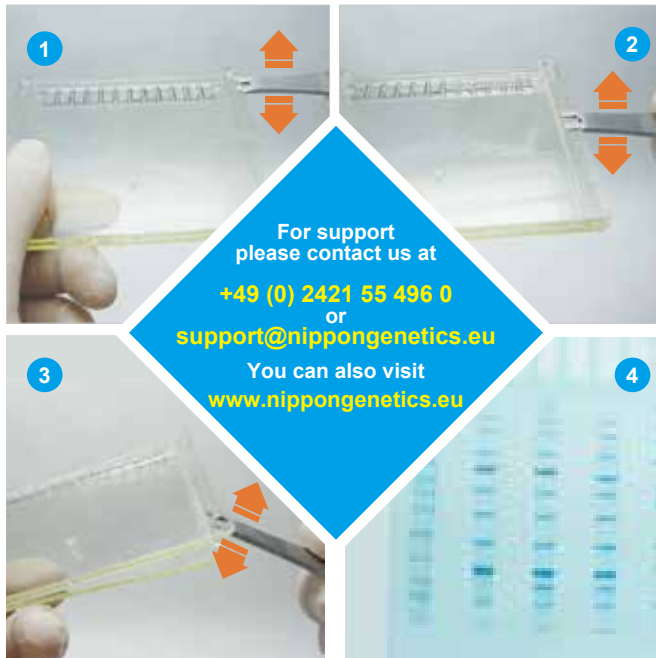
\*Running time is dependent on expected protein sizes, gel percentage and power supply used.

### 7. Compatibility

8 × 10 cm	10 × 10 cm
<ul style="list-style-type: none"> <li>- FastGene® Protein Chamber (NG-002)</li> <li>- Bio-Rad Mini-PROTEAN® II &amp; 3 &amp; Tetra System;</li> <li>- Hoefer SE250 Mighty Small II Mini &amp; SE260 Mighty Small II Deluxe</li> </ul>	<ul style="list-style-type: none"> <li>- LONZA PAGEr™ Minigel Chamber,</li> <li>- Hoefer SE260 Mighty Small II Deluxe,</li> <li>- Life Technologies Novex XCell Surelock® &amp; Bolt™ Mini Gel Tank*</li> </ul>

\*Extra cushion needed

### 8. Opening a gel cassette with an FastGene® Opener



# QuickGuide for FastGene® PAGE Gels 8 × 10 cm

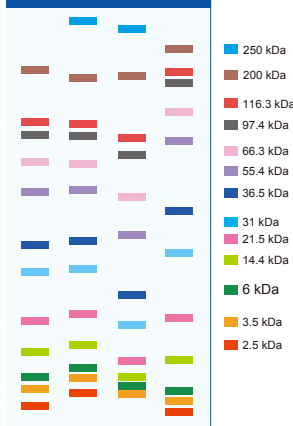


**NIPPON Genetics EUROPE**

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**1. Choose the appropriate gels for your protein electrophoresis analysis using the charts given below.**

8-16% 4-20% 4-12% 12%



Percentage	Separation range
8-16 %	10 kDa to 200 kDa
4-20 %	10 kDa to 250 kDa
4-12 %	20 kDa to 250 kDa
12 %	6.5 kDa to 200 kDa

**2. Preparation of sample:**

Reagent	Volume
Protein Sample*	x µl
Deionized H <sub>2</sub> O	Up to 8 µl
5 x Loading buffer	2 µl
<b>Total volume**</b>	<b>10 µl</b>

\*Heat the sample at 100 °C for 10 min (not for native gels).  
\*\*Maximal volume per well is 60 µl.

**3. Prepare the gel tank and the running buffer. Please use FastGene® MOPS (PG-MOPS10) or MES buffer as a PAGE running buffer. Do not use Tris-Glycine.**

**NOTE:** The FastGene® MOPS (PG-MOPS10) buffer contains SDS and is therefore not suitable for native PAGE.

MOPS Buffer	
Tris-base	6.06 g
MOPS	10.46 g
EDTA	0.3 g
(SDS)*	(1 g)*
H <sub>2</sub> O	up to 1000 ml

\*Not for native gels

MES Buffer	
Tris-base	6.06 g
MES	9.76 g
EDTA	0.3 g
(SDS)*	(1 g)*
H <sub>2</sub> O	up to 1000 ml

\*Not for native gels

**4. Remove the tape at the bottom of the gel plate and the comb gently, then insert the gel into the gel running apparatus and add the running buffer.**



Remove comb and tape

**NOTE:** For Bio-Rad tanks, turn around the gasket to fit the gel plate



Wrong!



Correct!

